

Full Length Research Paper

Bioinformatics techniques for designing antiviral agents against influenza

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Influenza virus infection is responsible for hundreds of thousands of deaths annually. Current vaccination strategies and antiviral drugs provide limited protection; therefore new strategies are needed. RNA interference is an effective means and is now widely used to knock-down gene expression in a sequence specific manner making it a powerful tool not only for studying gene function, but also for therapeutic purposes, including antiviral treatment, by targeting highly divergent pathogens including human immuno-deficiency virus (HIV), hepatitis C virus (HCV), influenza virus and SARS coronavirus, all of which pose enormous threats to global human health. In the present work, potential siRNA are being designed against segment HA and NA of influenza virus using a novel web-based online software system, siVirus, which provides functional, off-target minimized siRNAs targeting highly conserved regions of divergent viral sequences.

Keywords: influenza, antiviral agents, virus

INTRODUCTION

Influenza, commonly known as flu, is an infectious disease of birds and mammals caused by an RNA virus of the family *Orthomyxoviridae* (the influenza viruses). Also known as flu which is a contagious disease that is caused by the influenza virus. It attacks the respiratory tract in human (Carolyn *et al.*, 2003). Acute viral infection of the respiratory tract, occurring in isolated cases, in epidemics, or in pandemics; is caused by serologically different strains of influenza viruses designated as A, B, and C, has a 3-day incubation period, and usually lasts for 3 to 10 days (George E *et al.*, 2001). There are three genera of influenza virus, identified by antigenic differences in their nucleoprotein and matrix protein:

- *Influenzavirus A* are the cause of all flu pandemics and are known to infect humans, other mammals and birds (avian influenza).
- *Influenzavirus B* are known to infect humans and seals.
- *Influenzavirus C* are known to infect humans and pigs.

The type A viruses are the most virulent human pathogens among the three influenza types and causes

the most severe disease (Hay *et al.*, 2001). Influenza A viruses are further divided into subtypes based on the surface glycoproteins, hemagglutinin (HA) and neuraminidase (NA). At present, 16 HA subtype (H1–H16) and nine NA subtypes (N1–N9) have been recognized. Each virus has one HA and one NA antigen, apparently in any combination. (Suzuki., 2005). Influenza virus mutations are subtyped specific. siRNAs have a well-defined structure: a short (usually 21-nucleotide-long) double-strand of RNA (dsRNA) with 2-nucleotide 3' overhangs on either end. Each strand has a 5' phosphate group and a 3' hydroxyl (-OH) group. siRNAs can also be introduced into cells using nanoparticles and other methods to bring about the specific knockdown of a gene of interest. Essentially any gene of whose the sequence is known can thus be targeted based on sequence complementarity with an appropriately tailored siRNA. This method has made siRNAs an important tool for gene function and drug target validation studies in the post-genomic era (Qing G *et al.*, 2004). Bioinformatics approaches are significant to come up with intelligent solutions to existing health related issues e.g. designing molecules against virus infection. siRNA's can be used for gene silencing or for therapeutic purposes like vaccines (David B *et al.*, 2006).

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MATERIAL AND METHODS

Sequence data for gene segments 4 (hemagglutinin) and 6 (neuraminidase) of influenza A virus were downloaded from GenBank. To facilitate the computational analysis of these background datasets, those isolates with identical sequences were removed from the analysis. siVirus interface (<http://sivirus.rnai.jp/.html>) was used to obtain results. siVirus especially focuses on antiviral siRNA design and provides:

- Highly conserved target sites for designing antiviral siRNAs that would resist viral mutational escapes. siRNA sequences are selected based on their *degree of conservation*, defined as the proportion of viral sequences that are targeted by the corresponding siRNA, with complete matches (*i.e.*, 21/21 matches). All possible siRNA candidates targeting every other position of user-selected viral sequences are generated and their degrees of conservation are compared. Users can arbitrarily specify a set of viral sequences for the computation; for example, sequences can be selected from a specific geographic region(s) or a specific genotype(s) to design the best siRNAs tailored to specific user needs.

- Selection of Effective siRNAs based on guidelines of Ui-Tei *et al.*, Reynolds *et al.*, and Amarzguoui *et al.*

Those guidelines are:

1. A/U at the 5' end of the antisense strand
2. G/C at the 5' end of the sense strand
3. At least 5 A/U residues in the 5' terminal, one third of the antisense strand
4. The absence of any GC stretch of more than 9 nucleotide in length

If any of the above four guideline is not fulfilled then there is no gene silencing by a particular siRNA (Amarzguoui *et al.*, 2004).

Off-target minimized siRNAs. siVirus shows the number of off-target hits within two mismatches against human genes. Off-target hits are those which have dissimilarity to the target sequence. It is desirable to select a siRNA that has less off-target hits.

RESULTS

siRNA results with siVIRUS

A quite large number of possible siRNA candidates (*i.e.* substrings of length 21) targeting every other position of the Influenza virus A sequences were produced from the retrieved viral sequences. The analysis of these siRNA candidates revealed that highly conserved siRNAs constituted only 0.0% of the possible siRNAs if >90% conservation is expected. But the fraction increases to 100% in case of Hemmagglutinin segment of Influenza A

virus even if the threshold of the conservation is relaxed to 80%. On the other hand, siRNAs predicted to be functional by one or more guidelines as mentioned in material and methods constituted in case of all the HA sequences present in siVirus interface 65.35%, in case of region specific HA sequences 51.16%, in case of all the NA sequences 84.08% and in case of region specific NA sequences 78.78% of the siRNAs (Figure 1, 2, 3, and 4). Taken together, siRNAs showing maximum conservation are given in their duplex form. In this condition, siRNAs can be designed for each full-length sequence of segment HA and NA of Influenza virus A. Graphs in figure 1, 2, 3 and 4 shows probability of siRNAs fulfilling at least one guideline as mentioned in material and methods.

siRNA results with siDIRECT

siRNA duplex shows the double stranded sequence that is sense and anti-sense strands of siRNA. Oligo sequences means short sequences as here the length of sequence is 21bp. *Mismatch tolerance* describes the minimum number of mismatches between the siRNA sequence and any non-targeted sequence. An siRNA sequence of mismatch tolerance three does not match any off-target candidates with fewer than three mismatches. A higher mismatch tolerance of siRNA sequence indicates its high specificity in the presence of some mismatches.

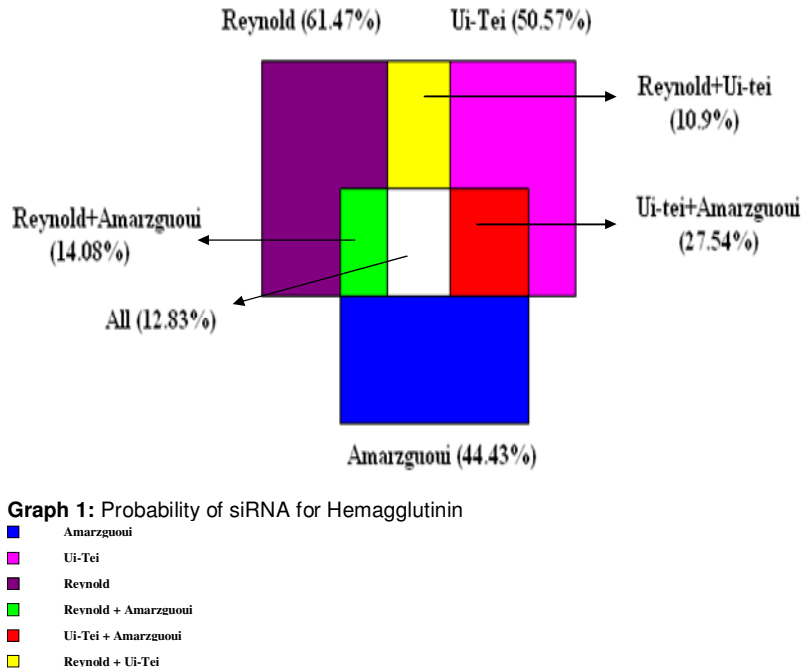
DISCUSSION

Influenza virus infection is a major public health problem, causing millions of cases of severe illness and as many as 500,000 deaths each year world wide. Factors contributing to the pathogenicity of influenza in human are not fully understood. Clearly the ability of the influenza virus to evolve rapidly and reassort contributes to the potential for epidemics and pandemics and may also contribute to the severity of disease. The restriction of influenza to the respiratory tract in human differentiates the disease from that in bird, in which replication is normally restricted to the GI tract but pathogenic strains can cause systemic infections. The HA and NA of the avian viruses has a crucial role in pathogenicity (Taubenberger *et al.*, 2000).

In start the scourge of Influenza has probably affected mankind for several millenia and is feared because of its dramatic impact during pandemics (Micheal *et al.*, 2001). Over the past few years novel class of antiviral agents, the neuraminidase inhibitors has been found to be safe and effective in the prevention and treatment of influenza. But they too provide partial protection. Previously available agents, the M2 inhibitors amantadine and rimantadine, could only be used to treat

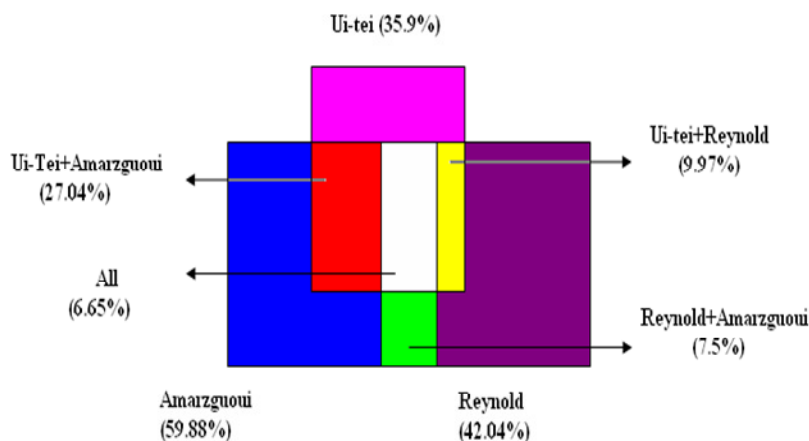
siRNA target site		siRNA efficacy prediction			≤ 2 mismatch	Conservation in the selected sequences
21bp target + 2nt overhang	map to segment 4	Ui-Tei	Reynolds	Amarzguoui	off-target hits	(+: conserved, -: not conserved, -: sequence not available)
AGCAACTGTTACCCCTTATGAtg		yes	yes		0 [detail]	83% (835/1006)
CAGCAACTGTTACCCCTTATGAtg		yes	yes	yes	0 [detail]	82.9% (834/1006)
TTCAAAATATACCACAAATGtg		yes			7 [detail]	82.3% (828/1006)
AGAATTTCAGGACCTCGAGAAAta			yes		1 [detail]	82.2% (827/1006)
AATTCAGGACCTCGAGAAATAtg			yes		3 [detail]	82.2% (827/1006)
TGGATTTCCTTCATATCAAtg		yes			1 [detail]	82.2% (827/1006)
GAATGCATTCCTTCATGGAag					1 [detail]	82.2% (827/1006)
TTGGTACGGTTTCAGGCATCAaa				yes	1 [detail]	81.9% (824/1006)
TGGTACGGTTTCAGGCATCAaa		yes	yes		1 [detail]	81.8% (823/1006)
GGTACGGTTTCAGGCATCAaa			yes		1 [detail]	81.8% (823/1006)
GTACGGTTTCAGGCATCAaa			yes	yes	1 [detail]	81.8% (823/1006)
TTTCAGGCATCAAAATCTGAgg			yes		18 [detail]	81.7% (822/1006)
AAGCATCTATTGGACAATAGTaa		yes	yes	yes	1 [detail]	81.6% (821/1006)
TGGGGTTTCATCATGTGGGCTGgc				yes	4 [detail]	81.5% (820/1006)
CTCCAAATGGGAAGCATTCCAAt		yes			7 [detail]	81.4% (819/1006)
TCCAAATGGGAAGCATTCCAAtg		yes			1 [detail]	81.4% (819/1006)
ATCACTCCAAATGGGAAGCATTCc		yes		yes	2 [detail]	81.3% (818/1006)
TACGGTTTCAGGCATCAAAATtc		yes	yes	yes	3 [detail]	81.2% (817/1006)
ACGGTTTCAGGCATCAAAATTct		yes	yes	yes	3 [detail]	81.2% (817/1006)
AAATGGAAGCATTCCTCAATGaca			yes		2 [detail]	80.8% (813/1006)
ATGGAAGCATTCCTCAATGACAaa		yes	yes	yes	3 [detail]	80.8% (813/1006)
TGGAAGCATTCCTCAATGACAAac		yes	yes	yes	1 [detail]	80.8% (813/1006)
AGGGAGAATTTCAGGACCTCGAga				yes	0 [detail]	80.7% (812/1006)
ATCATTGCTTTGAGCTACATtt		yes		yes	1 [detail]	80.6% (811/1006)
GAGAATTCAGGACCTCGAGAAat		yes	yes		0 [detail]	80.4% (809/1006)
GTGGGCTGCCCCAAAAGGCCAaca		yes		yes	3 [detail]	80.3% (808/1006)
TGGGCTGCCCCAAAAGGCCAacat				yes	2 [detail]	80.3% (808/1006)
GGGCTGCCCCAAAAGGCCAACatt				yes	0 [detail]	80.3% (808/1006)
GGCCTGCCCCAAAAGGCCAACAta		yes		yes	0 [detail]	80.3% (808/1006)
AGGCAACATTAGGTGCAACATtt		yes		yes	2 [detail]	80.2% (807/1006)

Figure 1: siRNAs for Hemagglutinin
A = siRNA sequence.
B = siRNA position.
C = siRNA efficacy predictions.
D = Off-target search results.
E = Degrees of conservation.
F = Conservation in the user-selected sequences.



siRNA target site		siRNA efficacy prediction			≤ 2 mismatch	Conservation in the selected sequences	
21bp target + 2nt overhang	map to segment 4	Ui-Tei	Reynolds	Amarzouioui	off-target hits	(+ : conserved, - : not conserved, - : sequence not available)	
CAGCAACTGTTACCCCTATGAtg		yes	yes	yes	0 [detail]	72.2% (86/119)	
AGCAACTGTATACCCCTATGAtgt		yes	yes	yes	0 [detail]	72.2% (86/119)	
TGGTACGGTTTCAGGCATCAAAa		yes	yes	yes	1 [detail]	72.2% (86/119)	
GGTACGGTTTCAGGCATCAAAat			yes		1 [detail]	72.2% (86/119)	
GTACGGTTTCAGGCATCAAAAt			yes	yes	1 [detail]	72.2% (86/119)	
TACGGTTTCAGGCATCAAAAttc			yes	yes	3 [detail]	72.2% (86/119)	
ACGGTTTCAGGCATCAAAAtct			yes	yes	3 [detail]	72.2% (86/119)	
TTTCAGGCATCAAAAtCTGAgg			yes	yes	18 [detail]	72.2% (86/119)	
ATCATTCGTTTGAGCTACATtt		yes		yes	1 [detail]	71.4% (85/119)	
CTTCAGAAGTAGAAGGGAGaAt				yes	7 [detail]	71.4% (85/119)	
CTCAGAAGTAGAAGGGAGAAItc		yes		yes	4 [detail]	71.4% (85/119)	
TGGATTCTCTTGCCATATCAtg		yes	yes		1 [detail]	71.4% (85/119)	
TATGACCATGATGTATACAGAgA			yes		4 [detail]	71.4% (85/119)	
TGACCATTGATGTATACAGAGAtg			yes		2 [detail]	71.4% (85/119)	
AGGGAGAATTACGACCTCGAgA				yes	0 [detail]	69.7% (83/119)	
AGAATTACGAGACCTCGAGAAAta			yes		1 [detail]	69.7% (83/119)	
AATTACGAGACCTCGAGAAAtAtg			yes		3 [detail]	69.7% (83/119)	
GGCAACATTAGGTGCAACATTtg		yes		yes	1 [detail]	69.7% (83/119)	
GAATGCATCACTCAAATGGAag			yes		1 [detail]	69.7% (83/119)	
GAGAATTACGAGACCTCGAGAAat		yes	yes		0 [detail]	68.9% (82/119)	
AGGCAACATTAGGTGCAACATtt		yes		yes	2 [detail]	68.9% (82/119)	
TTAGGTGCAACATTTCGATTtga				yes	2 [detail]	68.9% (82/119)	
TCGTAATGCATCACTCCAAATgg			yes		3 [detail]	68.9% (82/119)	
CTCAGAAATGGAGCAATTCcCAat		yes			7 [detail]	68% (81/119)	
TCCAAATGGGAAGCATTCcCAAtg		yes			1 [detail]	68% (81/119)	
AAATGGGAAGCATTCcCAATGAcA			yes		2 [detail]	68% (81/119)	
TCCAAAAATATCCACCAAAATgtGa			yes		7 [detail]	68% (81/119)	
AGGTGCAACATTTCGATTtGagt		yes	yes	yes	2 [detail]	68% (81/119)	
ATGGAAGCATTCcCAATGACAAac		yes	yes	yes	3 [detail]	67.2% (80/119)	
TGGGAAGCATTCcCAATGACAAac		yes	yes	yes	1 [detail]	67.2% (80/119)	

Figure 2: siRNAs for Asian and Russian Hemagglutinin



Graph 2: Probability of siRNA for Asian and Russian Hemagglutinin

Graph 2: Probability of siRNA for Asian and Russian Hemagglutinin

siRNA results with siDIRECT for Hemagglutinin

1. siRNA Information for 83% conservation:

siRNA duplex

5'-CAA CUG UUA CCC UUA UGA UGU sense strand

UUCGUUGACA AUGGGAAUACUA-5' antisense strand

Oligo sequences

>Sense_strand
CAACUGUUACCCUUAUGAUGU

>Antisense_strand
AUCAUAAGGGUACAGUUGCU

Mismatch tolerance

Both-strand mismatch tolerance: 3 Plus-strand mismatch tolerance: 3

2. siRNA Information for 82.9% conservation:

siRNA duplex

sRNA duplex

5'-GCAACUGUUACCCUUAUGAUG	sense strand
GUCGUUGACAAUGGGAAUACU-5'	antisense strand

Oligo sequences

```
>Sense_strand
GCAACUGUUACCCUUAUGAUG
>Antisense_strand
UCAUAAGGGGUAACAGUUGCUG
```

Mismatch tolerance

Both-strand mismatch tolerance:
Plus-strand mismatch tolerance: 3

3. siRNA Information for 82.3% conservation:

siRNA duplex

5'-CAAAUUAUACCACAAUGUGA sense strand
|||||
AAGUUUUUAUUGGUGUUUACA-5' antisense strand

Oligo sequences

```
>Sense_strand
CAAAAUAUACCACAAUGUGA
>Antisense_strand
ACAUUUGUGGUAAUUAUUUUGAA
```

Mismatch tolerance

Both-strand mismatch tolerance: 2 Plus-strand mismatch tolerance: 2

4. siRNA Information for 82.2% conservation:

i. siRNA duplex

5'-AAUUCAGGACCUCGAGAAUA sense strand
 |||
 UCUUAAGUCCUGGAGCUCUUU-5' antisense strand

Oligo sequences

```
>Sense_strand
AAUUCAGGACCUCGAGAAAUA
>Antisense_strand
UUUUCUGGAGGUUCGUGAAUUCU
```

Mismatch tolerance

Both-strand mismatch tolerance: 3 Plus-strand mismatch tolerance: 3

ii. siRNA duplex

5'-UUCAGGACCUCGAGAAUAUG sense strand
 |||||
UUAAGUCCUGGAGCUCUUUAU-5' antisense strand

Oligo sequences

```
>Sense_strand
UUCAGGACCUCGAGAAUAUG
>antisense_strand
UAUUUCUCGAGGUCCUGAAUU
```

Mismatch tolerance

Both-strand mismatch tolerance: 2 Plus-strand mismatch tolerance: 2

iii. siRNA duplex

5'-GAUUUCCUUUGCCAUUCAUG sense strand
 |||||
 ACCUAAAAGGAAACGGUUAUGU-5' antisense strand

Oligo sequences

```
>Sense_strand
GAUUUCCUUUGCCAUAUCAUG
>antisense_strand
UGAUAUGGCAAAGGAAAUCCA
```

Mismatch tolerance

Both-strand mismatch tolerance:3 Plus-strand mismatch tolerance: 3

5. siRNA Information for 82% conservation:

siRNA duplex

5'-AUGCAUCACUCCAAAUGGAAG sense strand
|||||
CUUACGUAGUGAGGUUUACCU-5' antisense strand

Oligo sequences

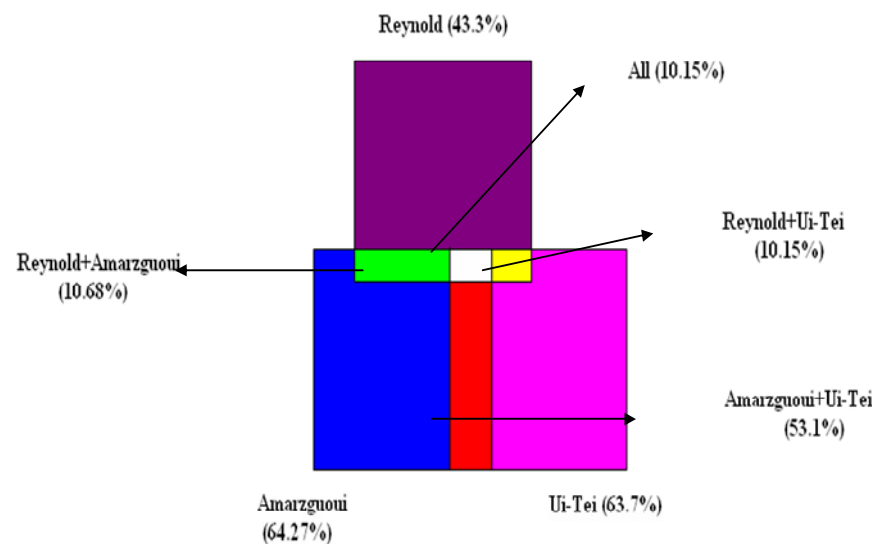
```
>Sense_strand
AUGCAUCACUCCAAAUGGAAG
>antisense_strand
UCCAUUUGGAGUGAUGCAUUC
```

Mismatch tolerance

Both-strand mismatch tolerance: 3 Plus-strand mismatch tolerance: 3

siRNA target site	map to segment 6	siRNA efficacy prediction	Ui-Tei	Reynolds	Amarzguoui	≤ 2 mismatch off-target hits	Conservation in the selected sequences
21bp target + 2nt overhang							(+: conserved, -: not conserved, -: sequence not available)
GTGGACCTCAACAGTATTGtg		yes			yes	0 [detail]	85.9% (1123/1307)
TGGACCTCAACAGTATTGtg		yes			yes	1 [detail]	85.8% (1122/1307)
GACCTCAACAGTATTGtg		yes			yes	2 [detail]	85.2% (1114/1307)
TAGCATGGTCCAGCTCAAGTtg		yes	yes	yes	yes	2 [detail]	85% (1112/1307)
ATGGTCCAGCTCAAGTTGTCAcg		yes			yes	4 [detail]	85% (1112/1307)
CTCAACAGTATTGTTGTGTTt		yes			yes	2 [detail]	84.9% (1110/1307)
TCCAGCTCAAGTTGTCACGATgg		yes			yes	1 [detail]	84.3% (1103/1307)
AACAGTATTGTTGTTTGTgg		yes			yes	9 [detail]	84.3% (1103/1307)
TCAACAGTATTGTTGTTTtg			yes			6 [detail]	84.3% (1102/1307)
CTCAAGTTGTCACGATGGAAAag		yes	yes	yes	yes	0 [detail]	84.1% (1100/1307)
AGCTCAAGTTGTCACGATGGAAA			yes			2 [detail]	84% (1099/1307)
TCAAGTTGTCACGATGGAAAagc			yes			0 [detail]	84% (1099/1307)
TGCAACTGCTAGCTTCATTTatg		yes	yes	yes	yes	0 [detail]	83.3% (1090/1307)
AATGCAACTGCTAGCTTCATTTa			yes			3 [detail]	83.3% (1089/1307)
ATGCAACTGCTAGCTTCATTTat		yes	yes	yes	yes	0 [detail]	83.3% (1089/1307)
AGTTGTCAAGTGGAAAAGCAtg				yes		2 [detail]	82.6% (1080/1307)
ATGACGTGTGGATGGGAAGAAca		yes			yes	2 [detail]	82.3% (1076/1307)
GAACCTTATGTGTCATGCGATcc					yes	0 [detail]	82.1% (1074/1307)
TGCTTTTATGTGGAGTTGATAag		yes			yes	1 [detail]	82.1% (1074/1307)
ATGTGGAGTTGATAAGGGGAagg					yes	1 [detail]	82.1% (1074/1307)
GTGCTTTTATGTGGAGTTGATAa			yes	yes	yes	2 [detail]	82% (1073/1307)
AATCCAAATCAAAAGATAATAac					yes	5 [detail]	81.5% (1066/1307)
ATCCAAATCAAAAGATAATAaca		yes	yes	yes	yes	6 [detail]	81.4% (1064/1307)
GAAAAGCATGGCTGCATGTTtg				yes		6 [detail]	81.3% (1063/1307)
TGGAAGCATGGCTGCATGTTt		yes				1 [detail]	81.2% (1062/1307)
ATTACAGGATTTGCACCTTTTtc			yes			2 [detail]	81.1% (1061/1307)
TTACAGGATTTGCACCTTTTct				yes	yes	1 [detail]	81.1% (1060/1307)
ACGTGTGGATGGGAAGAACGAtc		yes				0 [detail]	80.8% (1057/1307)
AACATAACAGAGATAGTGATtt		yes			yes	4 [detail]	80.7% (1056/1307)
ATTTGCACCTTTTCTAAGGAca			yes			3 [detail]	80.2% (1049/1307)

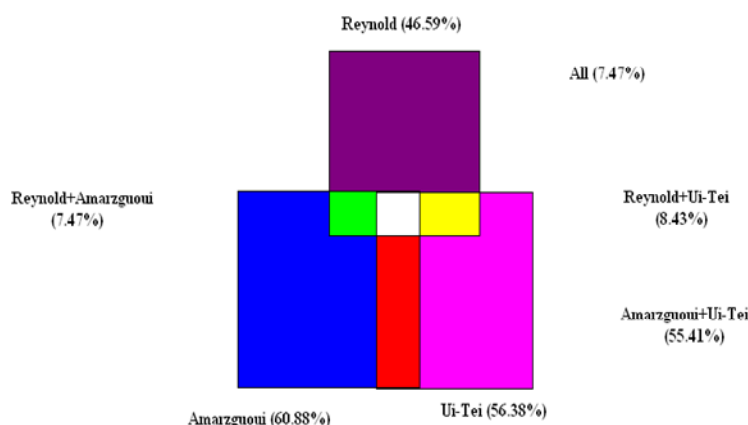
Figure 3: siRNAs for Neuraminidase



Graph 3. Probability of siRNA for Neuraminidase

siRNA target site		siRNA efficacy prediction		≤ 2 mismatch	Conservation in the selected sequences	
21bp target + 2nt overhang	map to segment 6	Ui-Tei	Reynolds	Amarguioui	off-target hits	(.:conserved, .:not conserved, .:sequence not available)
AATCAAATCAAAAGATAATAc			yes	yes	5 [detail]	75.3% (122/162)
ATGAATCCAAATCAAAAGATAa		yes	yes	yes	8 [detail]	74% (120/162)
GAATCCAAATCAAAAGATAATaa				yes	6 [detail]	74% (120/162)
ATCCAAATCAAAAGATAATAcA		yes	yes	yes	6 [detail]	74% (120/162)
TCAAACAGTATTGTGTGTTtg			yes		6 [detail]	74% (120/162)
AACAGTATTGTGTGTTTGTg		yes		yes	9 [detail]	74% (120/162)
ATGTCACAGCTCAAGTTGTCAc		yes		yes	4 [detail]	73.4% (119/162)
TCCAGCTCAAGTTGTGACAGTg			yes	yes	1 [detail]	73.4% (119/162)
GTGACCTCAAAACAGTATTGTg		yes		yes	0 [detail]	73.4% (119/162)
TGGACCTCAAAACAGTATTGTg		yes		yes	1 [detail]	73.4% (119/162)
GACCTCAAAACAGTATTGTGt		yes		yes	2 [detail]	73.4% (119/162)
CTCAAAACAGTATTGTGTGTTt		yes		yes	2 [detail]	73.4% (119/162)
TAGCATGTGCTCAAGTCAAGTgt			yes	yes	2 [detail]	72.8% (118/162)
GGAGTGAAGGCTGGGCCCTTga				yes	2 [detail]	72.8% (118/162)
CTCAAGTTGTACGATGGAAAG		yes	yes	yes	0 [detail]	72.2% (117/162)
TCAAGTTGTACGATGGAAAGc				yes	0 [detail]	72.2% (117/162)
AGTTGTACGATGGAAAGCAtg		yes		yes	2 [detail]	72.2% (117/162)
AGTGAAGGCTGGGCCCTTGAc			yes		4 [detail]	72.2% (117/162)
ATGAATGAGTTGGGTTGCCAt		yes	yes		1 [detail]	71.6% (116/162)
AATGAGTTGGGTTGCCATTCa			yes		2 [detail]	71.6% (116/162)
TGAGTTGGGTTGCCATTTCa			yes		1 [detail]	71.6% (116/162)
AGCTCAAGTTGTACGATGGAAa		yes		yes	2 [detail]	71.6% (116/162)
TGCAACTGCTAGCTTCATTAtg		yes	yes	yes	0 [detail]	71.6% (116/162)
AACATAACAGAGATAGTGTATt		yes		yes	4 [detail]	70.9% (115/162)
ATTGCACTTTTCTAAGGAAc			yes		3 [detail]	70.9% (115/162)
AATGCAACTGCTAGCTTCATTa			yes		3 [detail]	70.9% (115/162)
ATGCAACTGCTAGCTTCATTAt		yes	yes	yes	0 [detail]	70.9% (115/162)
TTGCACCTTTTCTAAGGACAAt		yes	yes	yes	3 [detail]	70.3% (114/162)
TGCACCTTTTCTAAGGACAAtc		yes		yes	2 [detail]	70.3% (114/162)
GCACCTTTTCTAAGGACAAtc				yes	2 [detail]	70.3% (114/162)

Figure 4: siRNAs for Asian and Russian Neuraminidase



Graph 4: Probability of siRNA for Asian and Russian Neuraminidase

➤ **siRNA results with siDIRECT for Neuraminidase**

1. siRNA Information for 85.9% conservation:

siRNA duplex

5'-GGACCUC AACAGUAUUGUUG sense strand
 |||||
 CACCUGGAGUUUGUCAUAACA-5' antisense strand

Oligo sequences

>Sense_strand

GGACCUCAAACAGUAUUGUUG

>Antisense_strand

ACAAUACUGUUUGAGGUCCAC

Mismatch tolerance

Both-strand mismatch tolerance: 3 Plus-strand mismatch tolerance: 3

2. siRNA Information for 85.8% conservation:**siRNA duplex**

```

5'-GACCUCAAACAGUAUUGUUGU      sense strand
   ||| ||| ||| ||| ||| ||| |||
ACCUGGAGUUUGUCAUAACAA-5'      antisense strand

```

Oligo sequences

```

>Sense_strand
GACCUCAAACAGUAUUGUUGU
>Antisense_strand
AACAAUACUGUUUGAGGUCCA

```

Mismatch tolerance

Both-strand mismatch tolerance: 3 Plus-strand mismatch tolerance: 3

2. siRNA Information for 85.2% conservation:**siRNA duplex**

```

5'-CCUCAAAACAGUAUUGUUGUGU      sense strand
   ||| ||| ||| ||| ||| ||| |||
CUGGAGUUUGUCAUAACAACA-5'      antisense strand

```

Oligo sequences

```

>Sense_strand
CCUCAAAACAGUAUUGUUGUGU
>antisense_strand
ACAACAUAUACUGUUUGAGGUC

```

Mismatch tolerance

Both-strand mismatch tolerance: 2 Plus-strand mismatch tolerance: 3

4. siRNA Information for 85% conservation:**i. siRNA duplex**

```

5'-GCAUGGUCCAGCUCAAGUUGU      sense strand
   ||| ||| ||| ||| ||| ||| |||
AUCGUACCAGGUCGAGUUCAA-5'      antisense strand

```

Oligo sequences

```

>Sense_strand
GCAUGGUCCAGCUCAAGUUGU
>Antisense_strand
AACUUGAGCGUGGACCAUGCUA

```

Mismatch tolerance

Both-strand mismatch tolerance: 2 Plus-strand mismatch tolerance: 2

ii. siRNA duplex

```

5'-GGUCCAGCUCAAGUUGUCACG      sense strand
   ||| ||| ||| ||| ||| ||| |||
UACCAGGUCGAGUUAACAGU-5'      antisense strand

```

Oligo sequences

```

>Sense_strand
GGUCCAGCUCAAGUUGUCACG
>Antisense_strand
UGACAACUUGAGCUGGACCAU

```

Mismatch tolerance

Both-strand mismatch tolerance: 2 Plus-strand mismatch tolerance: 2

5. siRNA Information for 74.6% conservation (Asia and Russia):**siRNA duplex**

```

5'-UCCAAAUCAAAGAUAAUAAAC      sense strand
   ||| ||| ||| ||| ||| ||| |||
UUAGGUUUAGUUUUAUUAU-5'      antisense strand

```

Oligo sequences

```

>Sense_strand
UCCAAAUCAAAGAUAAUAAAC
>Antisense_strand
UAUUAUCUUUUGAUUUGGAUU

```

Mismatch tolerance

Both-strand mismatch tolerance: 2 Plus-strand mismatch tolerance: 2

Influenza A infection and resistance develops rapidly (Wright *et al.*, 2002).

Inactivated vaccines are 60-80% effective against matched influenza strains (Hannon, 2002). Vaccination coverage is a problem world wide. Moreover, this strategy provides no protection against unexpected strains, outbreaks such as H5 and H7 avian influenza outbreaks in Honk Kong in 1997 and the Netherlands and Southeast Asia in 2003—2004, over pandemics.

RNA interference is an effective means of suppressing virus replication. Small interfering RNAs (siRNAs),

mediators of RNAi, are short (21-26nt), double-stranded RNA duplexes that inhibit gene expression by inducing sequence-specific degradation of homologous mRNA (Gitlin *et al.*, 2003). A number of studies demonstrated inhibition of replication of RNA viruses by RNAi (Silva *et al.*, 2002) (jacque *et al.*, 2002), including HIV (Lee *et al.*, 2003), Polio virus (kapadia *et al.*, 2003), Hepatitis C virus (Randal *et al.*, 2003), West Nile virus, and influenza virus (Mcmanus *et al.*, 2003).

siRNA analysis through siVirus software has been done before for HIV in 495 near full-length HIV-1

sequences were analyzed. The analysis of those siRNA candidates revealed that highly conserved siRNAs constituted only 0.3% of the possible siRNAs for >90% conservation. The fraction was still as small as 0.8% even the threshold of the conservation was relaxed to 80%. On the other hand, siRNAs predicted to be functional by one or more guidelines constituted 35.5% of the 4417157 siRNAs. Taken together, siRNAs that were >80% conserved, and satisfied at least one guideline constituted only 0.2% of the siRNAs. In our results 0% highly conserved siRNAs were obtained while using >90% conservation parameter. However on decreasing the percentage of conservation close to 80% than siRNAs constituted 100% conservation in case of neuraminidase. We reported a screen of 90522 siRNA candidates for targeting 1006 sequences of the HA and 13576 siRNA candidates for targeting 1307 sequences of the NA segment of influenza genome.

Our results showing typical output from siVirus for designing anti-Influenza Virus A siRNAs. The sequences from Influenza Virus A subtypes A and B, which are the most prevalent Influenza Virus A genotypes circulating in Asia, were selected. The results were sorted by the degree of conservation, and filtered to display siRNAs that satisfy at least one efficacy guideline. It is desirable to select a siRNA that has less off target hits as they predict the dissimilarity with the target sequence. With 0 or 1 off-target hits we got 17 siRNA in case of Hemagglutinin and 13 siRNAs in case of Neuraminidase.

To test the validity of siVirus, siRNAs satisfying the guideline by Ui-Tei *et al.* showed better results i.e. the maximum conserved regions of Influenza Virus A segments HA and NA using siVirus. Thus guidelines of Ui-tei *et al* predicted efficient siRNA against influenza A virus.

CONCLUSION

The efforts to develop antiviral agents directly from viral genome sequences has led to short interfering RNA (siRNA), the mediator of a sequence-selective inhibition known as RNA interference (RNAi). The current *in silico* study resulted in finding various siRNA agents which can initially be tested in animal model and the results may have implications in developing prophylactic as well as therapeutic treatment against influenza infection. The outcome of this study supports the growing expectation that siRNA can fulfill the need for moving rapidly from gene sequence to targeted therapeutic agents for many previously intractable disease targets. siRNA can be effective when given to animals before or in established influenza infections and this intervention would be useful during influenza outbreaks. Further development of this technology may provide an effective strategy for controlling influenza and other viral diseases.

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